



## VETBIONET

Veterinary Biocontained facility Network for excellence in animal infectiology research and experimentation

### Deliverable D19.1

***Quantity of access provided over the duration of the project to IRTA Animal and BSL3 laboratories facilities***

**Due date of deliverable: M72**

**Actual submission date: M72**

**Start date of the project: March 1<sup>st</sup>, 2017                      Duration: 72 months**

**Organisation name of lead contractor: Your organisation**

**Revision: V1**

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|---|----------|
| <b>Dissemination level</b>  |          |
| <b>Public</b>   | <b>X</b> |
| <b>Confidential, only for members of the consortium (including Commission Services)</b> |          |
| <b>Classified, as referred to in Commission Decision 2001/844/EC</b>                    |          |

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## 1. TNA Provided

| Name of the TNA project  | Name of TNA user | Organisation of TNA user   | Country of TNA user | Installation from the RI                | Start date | End date   | Number of units of access provided |
|--|------------------|--|---------------------|---|------------|------------|------------------------------------|
| Experimental infection of ewes and pigs with a novel pestivirus closely related to classical swine fever virus | Ana Maria Moreno | Istituto Zooprofilattico Sperimentale della Lombardia ed Emilia Romagna - IZSLER | Italy               | Animal and BSL3 laboratories facilities | 19/07/2019 | 01/03/2020 | 1                                  |
| Comparison of follicular B-cell maturation during PRRSV and Influenza virus (IAV) infections in swine          | Nicolas Bertho   | Ecole Nationale Vétérinaire Agroalimentaire et de l'Alimentation-ONIRIS          | France              | Animal and BSL3 laboratories facilities | 14/09/2021 | 24/11/2021 | 1                                  |

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## 2. Final reports of each TNA provided

### 2.1 TNA 1

#### **Experimental infection of ewes and pigs with a novel pestivirus closely related to classical swine fever virus**

The aim of the project was to study in two different animal species, the host range, clinical disease and pathogenic role of a new pestivirus detected in 2017 in Italy. First, we investigated the effects of this pestivirus in its natural hosts (sheep), with particular attention to the abortogenic role of this pestivirus and therefore to the effects in pregnant ewes and its offspring. Secondly, we studied its ability to infect pigs and to induce disease. Finally, we investigated the ability of this virus to induce an immunological response in pigs capable of protecting animals against a challenge with a highly virulent classical swine fever virus (CSFV) strain, as well as the possible serological cross-reactions between the new pestivirus and CSFV.

The abortogenic role of the Italian pestivirus was demonstrated by the abortions, stillbirths or weak lamb births recorded in all inoculated ewes and by the high viral RNA load detected in all foetal tissues. This pestivirus also demonstrated the ability to infect pigs, but with mild clinical signs. Interestingly, pigs experimentally infected with the new pestivirus developed immunological responses capable of protecting them from infection with a highly virulent strain of bluetongue virus (challenge). These animals showed only mild CSF symptoms compared to the infected control group, and a low viral RNA load was detected in one pig compared to the high viral titres observed in all control pigs.

Finally, a close serological and genomic relationship between the new ovine pestivirus and CSFV was evidenced using serological and RNA detection methods. Further studies are needed to investigate the prevalence of this new pestivirus in sheep and pigs.

Ana Maria Moreno and Enrica Sozzi from IZSLER stayed 3 days at CReSA in December 2019 to discuss and analyse the results obtained from this TNA. Biological material was sent to IZSLER.

From the results provided by this TNA, 2 publications were produced: doi: 10.3390/v12070775 and doi: 10.1111/tbed.14119.

*NB: Due to the complexity and length of this TNA and the need for animal and laboratory BSL3 facilities, this work required 1 and 1.5 access units for the Animal facility and BSL3/2 laboratories, respectively (see D18.1 and D20.1).*

### 2.2 TNA 2

#### **Comparison of follicular B-cell maturation during PRRSV and Influenza virus (IAV) infections in swine**

Porcine reproductive and respiratory syndrome virus (PRRSV) infections are responsible of serious respiratory problems and their persistence in herds has resulted in an estimated cost

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of 1,503 million € per year in Europe. While porcine influenza infections last 1 week, PRRSV infections can persist for several weeks/months, hence the economic impact.

It is hypothesized that this persistence is due to a delay in the appearance of neutralizing antibodies, which should appear one week after infection, as is observed in influenza challenges. Neutralizing antibodies are produced through a maturation process of antibody-producing cells, B lymphocytes, within the lymph nodes (LN), but so far, no information has been published on how PRRSV interferes with this process. In order to clarify this point, we infected animals with influenza virus or PRRSV and compared the maturation process of B cells for 6 days post-infection. This required intensive use of the cytometer/cell sorter in CReSA BSL3 facilities.

Nicolas Bertho stayed at CReSA for the entire duration of the experiment, with the exception of one week corresponding to extension of acclimatisation of pigs (see below). Caroline Hervet from INRAE-ONIRIS stayed one week at BSL3 laboratories of CReSA to help sorting of the immune cells and conditioning them for further analysis.

The collected samples were sent to the project participants:

- Gaëlle SIMON and Olivier BOURRY (ANSES, Ploufragan) received the sera for neutralizing antibodies assays.
- Daniel DORY (ANSES, Ploufragan) received PBMC in RLT Buffer to analyse antibody diversity by CDR3 sequencing.
- Nicolas BERTHO received the frozen PBMC, the rest of the frozen lymph node cells as well as the whole lymph nodes in Tissuetek/OCT for further analysis of differentiation of blood and LN B cells upon PRRSV and swine influenza virus infection.

Comprehensive laboratory analyses were conducted in 2022 and extended into 2023, with additional experiments needed to complete the initial results.

*NB: Pigs were purchased in Andalusia from a farm that did not practice vaccination against PRRS or influenza virus. Upon arrival at the CReSA animal facilities, they presented mild respiratory symptoms. Viral and severe bacterial infections were ruled out after laboratory diagnostic. After an additional week of acclimatisation, pigs recovered a healthy status and infections could be performed according to the initial protocol. This additional week of acclimatisation involved the use of 0.375 AU for the Animal facility (see deliverable D18.1) in addition to the 1 AU initially required for Animal and BSL3 laboratories facilities (reported here). Additional 0.5 AU for BSL3/2 laboratories were also required to perform cytometry and cell sorting experiments on the various samples generated in this TNA project.*